

A Novel Xanthone from *Garcinia oligantha*

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One novel xanthone, oliganthone A (**1**), was isolated from the stems of the plant *Garcinia oligantha*. It features the O-bearing C(3)-atom and absence of C(4) compared with the structures of related known xanthenes, which have never been reported before. The structure of this compound was elucidated by spectroscopic analysis. Compound **1** showed strong HeLa cell growth-inhibiting effects with IC_{50} values below 10 μM .

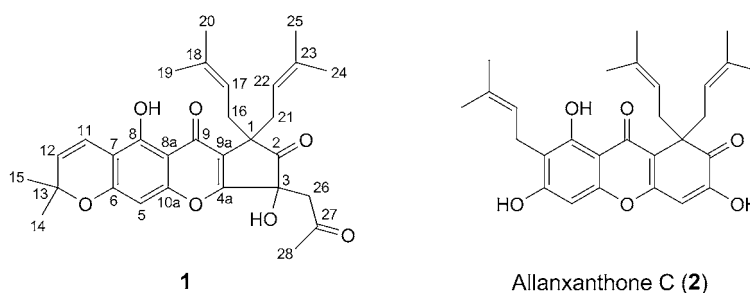
Introduction. – *Garcinia oligantha* (Guttiferae family), a 1–3-m-tall shrub, is distributed, at 200–1200 m altitude, in dense forests in Hainan and Guangdong Provinces [1]. We have already reported the isolation and characterization of various bioactive compounds including terpenes, xanthenes, benzophenones, and depsidones from *Garcinia paucinervis* [2][3], *G. multiflora* [4], and *G. oligantha* [5]. In our continuing search for biologically active and structurally unique compounds from the genus *Garcinia*, one novel polyisoprenylated xanthone, oliganthone A (**1**), together with known flavonoids, were isolated from the acetone extract of the dried stems *Garcinia oligantha*. Herein, we describe the isolation and structure elucidation of **1**. The compound was also assayed for cytotoxic activity against HeLa C3 cells.

Results and Discussion. – Oliganthone A (**1**), obtained as a yellow amorphous powder from an acetone extract of the stems of *G. oligantha*, exhibited a molecular-ion peak at m/z 507.2384 ($[M+H]^+$) in the HR-ESI-MS spectrum, corresponding to the molecular formula $\text{C}_{30}\text{H}_{34}\text{O}_7$, which indicated 14 degrees of unsaturation. The inspection of its ^1H - and ^{13}C -NMR data (*Table*) indicated the presence of seven Me, three CH_2 , and six CH (five olefinic) groups, and fourteen quaternary C-atoms (nine olefinic, one O-bearing, and three $\text{C}=\text{O}$). The absorption bands in its IR spectrum suggested the presence of OH groups (3447 cm^{-1}) and of a xanthone $\text{C}=\text{O}$ group (1656 cm^{-1}). With the aid of 1D- and 2D-NMR experiments, all ^1H - and ^{13}C -NMR signals of **1** were assigned by comparison of its NMR data with those of reference compounds. The deduced structure of **1** is shown in *Fig. 1*, and the ^1H - and ^{13}C -NMR data of **1** are compiled in the *Table*.

Table. ^1H - and ^{13}C -NMR Data of Compound **1** in CDCl_3^{a} . Arbitrary atom numbering as indicated in Fig. 1; δ in ppm, J in Hz.

Position	$\delta(\text{H})$	$\delta(\text{C})$	Position	$\delta(\text{H})$	$\delta(\text{C})$
1		58.5 (<i>s</i>)	15	1.81 (<i>s</i>)	28.2 (<i>q</i>)
2		212.2 (<i>s</i>)	16	2.60–2.66 (<i>m</i>)	33.6 (<i>t</i>)
3		75.9 (<i>s</i>)	17	4.89–4.91 (<i>m</i>)	119.7 (<i>d</i>)
5	6.30 (<i>s</i>)	100.7 (<i>d</i>)	18		135.9 (<i>s</i>)
6		162.0 (<i>s</i>)	19	1.56 (<i>s</i>)	17.7 (<i>q</i>)
7		101.7 (<i>s</i>)	20	1.64 (<i>s</i>)	25.9 (<i>q</i>)
8		159.7 (<i>s</i>)	21	2.70–2.75 (<i>m</i>)	34.6 (<i>t</i>)
9		180.1 (<i>s</i>)	22	4.85–4.89 (<i>m</i>)	118.9 (<i>d</i>)
4a		164.0 (<i>s</i>)	23		135.6 (<i>s</i>)
8a		106.0 (<i>s</i>)	24	1.53 (<i>s</i>)	17.7 (<i>q</i>)
9a		119.8 (<i>s</i>)	25	1.58 (<i>s</i>)	25.8 (<i>q</i>)
10a		152.2 (<i>s</i>)	26	2.77–2.82 (<i>m</i>)	42.8 (<i>t</i>)
11	6.63 (<i>d</i> , $J=10.0$)	114.4 (<i>d</i>)	27		208.4 (<i>s</i>)
12	5.61 (<i>d</i> , $J=10.0$)	127.8 (<i>d</i>)	28	2.26 (<i>s</i>)	31.9 (<i>q</i>)
13		78.3 (<i>s</i>)	HO–C(3)	5.15 (<i>br.</i>)	
14	1.49 (<i>s</i>)	28.4 (<i>q</i>)	HO–C(8)	12.66 (<i>s</i>)	

^a) Spectra were recorded with a Bruker DRX-400 MHz spectrometer, chemical shifts (δ) were expressed in ppm, J in Hz; assignments were confirmed by ^1H , ^1H -COSY, HMQC, and HMBC, experiments.

Fig. 1. Structures of compounds **1** and **2**

The ^1H -NMR spectrum of **1** displayed a set of signals at $\delta(\text{H})$ 4.89–4.91 and 4.85–4.89 (*m*, H–C(17) and H–C(22), resp.), 2.60–2.66 and 2.70–2.75 (*m*, CH_2 (16) and C(21), resp.), 1.56, 1.64, 1.53, and 1.58 (*s*, Me(19), Me(20), Me(24), and Me(25), resp.). These resonances are typical of a geminal bis(3-methylbut-2-en-1-yl) group, linked to a sp^3 -C-atom ($\delta(\text{C})$ 58.5 (C(1))) [6][7]. Except the signal of the xanthenone C=O group at $\delta(\text{C})$ 180.1 (C(9)), an additional C=O signal at $\delta(\text{C})$ 212.2 (C(2)) suggested a xanthenedione skeleton. The ^1H - and ^{13}C -NMR spectra of **1** displayed signals for a xanthenedione skeleton that contains three isoprene units [6]. Further comparison with the isolated compounds confirmed that **1** was an analog of a known xanthenedione named allanxanthone C [6].

Compared with the NMR data of allanxanthone **2** [6], this new compound did not exhibit signals for a third prenyl group. One of the C_5 group was established to be attached to C(6), forming a dimethyl-2*H*-pyrano-xanthone skeleton, by the HMBCs of $\delta(C)$ 162.0 (C(6)) with the chelated HO–C(8), and the *cis*-coupled H–C(11) and H–C(12), and those of H–C(12) with the O-bearing quaternary C(13) and the Me(14) and Me(15). The other two 3-methylbut-2-enyl groups are located at C(1), according to the HMBCs of H–C(16) with C(1), and of H–C(21) with C(2). The remaining aromatic H-atom was indicated to be H–C(5) by its HMBCs with C(6), C(7), C(8a), and C(10a) (Fig. 2).

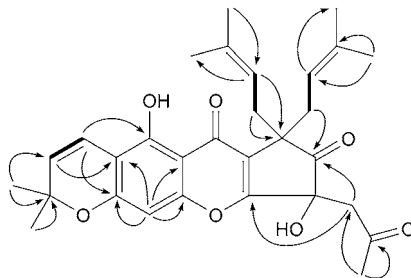


Fig. 2. Selected HMB (H \rightarrow C) and $^1H, ^1H$ -COSY (\rightleftharpoons) correlations of **1**

However, different C- and H-atom chemical shifts for C(3), C(26), C(27), and C(28) indicated that the structure of **1** differed from that of allanxanthone **2** [6] with respect to the side chain at C(3). In the HMBC spectrum of **1**, the correlations of the H-atom signals at $\delta(H)$ 5.15 (br., HO–C(3)) with the C-atom signals at $\delta(C)$ 212.2 (C(2)), 164.0 (C(4a)), and 42.8 (C(26)) suggested that C(4a) and C(3) linked to each other directly, which is without precedent. The O-bearing C-atom signal at $\delta(C)$ 75.9 (C(3)) and the absence of the C(4) signal further confirmed the above deduction. The O-bearing C(3)-atom and absent C(4) suggested compound **1** as a new xanthone, and shed a new insight into structural diversity of xanthone analogs. The HMBCs of the signals at $\delta(H)$ 2.77–2.82 (*m*, CH₂(26)) with those at $\delta(C)$ 212.2 (C(2)), 75.9 (C(3)), 208.4 (C(27)), and 31.9 (C(28)), and of the signals at $\delta(H)$ 2.26 (*s*, Me(28)) with those at $\delta(C)$ 42.8 (C(26)) and 208.4 (C(27)) indicated an acetyl moiety at C(3). Since we used acetone as solvent in the extraction procedure, we tested the acetone, MeOH, and CH₂Cl₂ extracts of the dried plant material by LC/MS, respectively. As a result, we detected the peak corresponding to compound **1** in all of the three extracts, which indicated that compound **1** was not an artefact of the extraction procedure. From these spectroscopic data, the structure of compound **1** was determined as shown, and it was named oliganthone A.

To determine the cytotoxicity of the compound, we measured its the IC_{50} value on HeLa cells. Oliganthone A (**1**) showed strong HeLa cell growth-inhibiting effects with the IC_{50} value of $7.27 \pm 0.23 \mu M$, which is below 10 μM .

Experimental Part

General. Column chromatography (CC): silica gel *GF*₂₅₄ (SiO₂; 200–300 mesh, *Qingdao Marine Chemical, Inc.*); *Sephadex LH-20* (*Pharmacia*); reversed-phase (RP) C₁₈ SiO₂ (250 mesh; *Merck*). TLC: Precoated SiO₂ 60 *GF*₂₅₄ (*Qingdao Marine Chemical, Inc.*). HPLC: *Agilent 1100* series, with an *Alltima C₁₈* column (4.6 × 250 mm) for HPLC analysis, and semi-prep. *Alltima C₁₈* columns (9.4 × 250 mm) for sample preparation. Optical rotation: *JASCO DIP-1000* polarimeter. UV Spectra: *Perkin-Elmer Lambda L14* spectrometer; λ_{\max} (log ϵ) in nm. IR Spectra: *Perkin-Elmer Spectrum 100* FT-IR spectrometer in KBr pellets; $\tilde{\nu}$ in cm⁻¹. ¹H- and ¹³C-NMR spectra (CDCl₃): *Bruker AV-400* spectrometer at 400 (¹H) and 100 MHz (¹³C); δ in ppm rel. to Me₄Si as internal standard, *J* in Hz. HR-ESI-MS: nanoLC-MS/MS system, with a *nanoAcquity UPLC* module and a *Q-TOF* spectrometer equipped with a nanoelectrospray ion source (*Waters*, Milford, MA; in *m/z*).

Plant Material. The stems of *G. oligantha* were collected in October 2008 in Hainan Province, P. R. China. The plant was identified by *Rong-Jing Zhang*. A voucher specimen (YNGXM-0001) has been deposited with the Key Laboratory of Ethnic Medicine Resource Chemistry (Yunnan University of Nationalities), State Ethnic Affairs Commission & Ministry of Education.

Extraction and Isolation. The air-dried and powdered stems of *G. oligantha* (5.5 kg) were extracted with acetone (5.0 l × 3, 24 h each) at r.t., and the extract was concentrated under vacuum condition. Then, the acetone extract was partitioned between H₂O and CH₂Cl₂. The CH₂Cl₂-soluble portion (242 g) was decolorized by *MCI GEL*[®] and eluted with 90% MeOH. The decolorized portion (130 g) was subjected to column chromatography (CC; SiO₂; hexane/acetone 1:0, 4:1, 2:1, 1:1, and 0:1) to afford five fractions, *Frs. I–V*. *Fr. I-1* (5.05 g) was then separated by CC (*RP-18*; MeOH/H₂O 80–100%) to give 14 fractions, *Frs. I-1.1–I-1.14*. *Fr. I-1.4* was separated by CC (*Sephadex LH-20*; MeOH) and then subjected to semi-prep. HPLC (MeOH/H₂O 80:20) to yield *oliganthone A* (**1**; 5.2 mg).

Bioassay. We determined the *IC*₅₀ values of **1** by using MTT (= 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-2H-tetrazolium bromide) assay. MTT Powder was dissolved in PBS (phosphate-buffered saline) at a concentration of 5 mg/ml. First, 2,500 HeLa cells suspended in 100 μ l of MEM (minimum essential *Eagle* medium) were seeded in a 96-well plate. After 24 h incubation, fresh medium containing various concentrations of each compound were added into the 96-well plate to replace the old medium. The concentrations applied were ranged from 100 to 1.56 μ M, which was achieved by twofold dilutions for 6 times. After 72-h treatment, 10 μ l of MTT soln. was added into each well of a 96-well plate. After 2 h incubation at 37°, 100 μ l of 10% SDS (sodium dodecyl sulfate) soln. with 0.01M HCl was added to dissolve the purple crystals. After 24 h incubation, the optical density (*OD*) values at 595 nm of the control groups at 0 and 72 h together with the compound-treated groups at 72 h from the MTT assay were measured using a plate reader. Paclitaxel at 500 nM was used as the positive control. *IC*₅₀ is the concentration of a compound inhibiting 50% of the cell growth.

Oliganthonone A (= 7,9-Dihydro-5,9-dihydroxy-2,2-dimethyl-7,7-bis(3-methylbut-2-en-1-yl)-9-(2-oxo-propyl)-2H-cyclopenta[e]benzo[1,2-b:5,4-b']dipyran-6,8-dione; **1**). Yellow amorphous powder. $[\alpha]_{\text{D}}^{25.3} = -3.6$ (*c* = 0.03, MeOH). UV (MeOH): 269 (4.08), 228 (2.68), 205 (2.34). IR (KBr): 3447, 3377, 3133, 2286, 1656, 1597, 1439, 1266, 1122, 1064, 861, 817, 621, 569. ¹H- and ¹³C-NMR: *Table*. HR-ESI-MS (pos.): 507.2384 ($[M + H]^+$, C₃₀H₃₅O₇⁺; calc. 507.2384).

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REFERENCES

- [1] X. W. Li, J. Li, P. F. Stevens, in 'Flora of China', Science Press, Beijing, 2007, Vol. 13, p. 45.
- [2] X. M. Gao, T. Yu, F. Lai, Y. Zhou, X. Liu, C. F. Qiao, J. Z. Song, S. L. Chen, K. Q. Luo, H. X. Xu, *Bioorg. Med. Chem.* **2010**, *18*, 4957.
- [3] X. M. Gao, T. Yu, F. Lai, C. F. Qiao, Y. Zhou, X. Liu, J. Z. Song, K. Q. Luo, H. X. Xu, *Tetrahedron Lett.* **2010**, *51*, 2442.

- [4] X. Liu, T. Yu, X. M. Gao, Y. Zhou, C. F. Qiao, Y. Peng, S. L. Chen, K. Q. Luo, H. X. Xu, *J. Nat. Prod.* **2010**, 73, 1355.
- [5] X. M. Gao, T. Yu, M. Z. Cui, J. X. Pu, X. Du, Q. B. Han, Q. F. Hu, T. C. Liu, K. Q. Luo, H. X. Xu, *Bioorg. Med. Chem. Lett.* **2012**, 22, 2350.
- [6] A. G. B. Azebaze, M. Meyer, A. Valentin, E. L. Nguemfo, Z. T. Fomum, A. E. Nkengfack, *Chem. Pharm. Bull.* **2006**, 54, 111.
- [7] M. Inuma, T. Ito, H. Tosa, T. Tanaka, R. Miyake, V. Chelladura, *Phytochemistry* **1997**, 46, 1423.

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